CATALOG DOCUMENTATION
NATIONAL COASTAL ASSESSMENT- NORTHEAST DATABASE
YEAR 2001 STATIONS
SEDIMENT CHEMISTRY DATA: "SEDCHEM"

TABLE OF CONTENTS

1. DATASET IDENTIFICATION

1.1 Title of Catalog document
National Coastal Assessment-Northeast Region Database
Year 2001 Stations
SEDIMENT CHEMISTRY DATA

1.2 Authors of the Catalog entry
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1.3 Catalog revision date
December 29, 2003

1.4 Dataset name
SEDCHEM

1.5 Task Group
National Coastal Assessment-Northeast

1.6 Dataset identification code
007

1.7 Version
001

1.8 Requested Acknowledgment
EMAP requests that all individuals who download EMAP data acknowledge the source of these data in any reports, papers, or presentations. If you publish these data, please include a statement similar to: "Some or all of the data described in this article were produced by the U. S. Environmental

Protection Agency through its Environmental Monitoring and Assessment Program (EMAP)".

2. INVESTIGATOR INFORMATION (for full addresses see Section 13)

2.1 Principal Investigators

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2.2 Sample Collection Investigators

Donald Cobb, U.S. EPA NHEERL-AED

2.3 Sample Processing Investigators

Not applicable

3. DATASET ABSTRACT

3.1 Abstract of the Dataset

The SEDCHEM data file reports the concentrations of chemical contaminants in sediment samples collected in Northeast estuaries sampled during the summer of 2001. Sediment samples were analyzed for 86 chemical constituents, including metals, polynuclear aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), and pesticides. One record is presented per analyte. For concentration values smaller than the MDL (non-detects), the result is reported as zero, the method detection limit (MDL) is listed, and the record is flagged; thereby giving the data user options for alternative treatment of non-detects (see Section 4.3). Note that consistent methods were not followed by all analytical laboratories in two respects: (1) in the digestion method used to analyze metals, and (2) in the MDL values used for most chemical analyses (see Section 5.2.6).

3.2 Keywords for the Dataset

Sediment contaminants, metals, polynuclear aromatic hydrocarbons, PAH, polychlorinated biphenyls, PCB, pesticides, DDT.

4. OBJECTIVES AND INTRODUCTION

4.1 Program Objective

The National Coastal Assessment (NCA) is a national monitoring and assessment program with the primary goal of providing a consistent evaluation of the estuarine condition in U.S. estuaries. It is an initiative of the Environmental Monitoring and Assessment Program (EMAP), and is a partnership of several federal and state environmental agencies, including: EPA's Regions, Office of Research and Development, and Office of Water; state environmental protection agencies in the 24 marine coastal states and Puerto Rico; and the United States Geological Survey (USGS) and the National Oceanic and Atmospheric Agency (NOAA). The five-year NCA program was initiated in 2000, and is also known as the Coastal 2000

Program.

Stations were randomly selected using EMAP's probabilistic sampling framework and were sampled once during a summer index period (June to October). A consistent suite of indicators was used to measure conditions in the water, sediment, and in benthic and fish communities. The measured data may be used by the states to meet their reporting requirements under the Clean Water Act, Section 305(b). The data will also be used to generate a series of national reports characterizing the condition of the Nation's estuaries.

4.2 Dataset Objective

A two-year sampling design was employed for 2000-2001 NCA program in the Northeast. Analysts may therefore wish to consider the two years of data together.

The objective of the sediment chemistry data file is to report the concentrations of chemical contaminants in estuarine sediment samples collected in the northeast NCA program in 2001.

4.3 Dataset Background Discussion

The suite of analytes measured are very similar to the contaminants measured by EPA's Environmental Monitoring and Assessment Program (EMAP) and NOAA's National Status and Trends program. Four classes of analytes are measured: polynuclear aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), organo-chlorine pesticides, and metals. Twenty-two PAHs are measured, consisting of the 16 priority pollutants defined by the Superfund program and several alkylated derivatives that prove to be useful in identifying sources of these compounds. The concentrations of 20 PCBs and 20 pesticides, all Superfund priority pollutants, are also measured. The NCA protocol calls for measuring total metal concentrations, which includes an aggressive HF/HNO3 digestion prior to analysis. As is discussed below, some samples received a less aggressive HNO3 digestion. Sediment grain-size and Total Organic Carbon (TOC) measurements made on the same sediments are reported in the SEDGRAIN file.

Routinely, the concentration values from clean sites were reported as smaller than the method detection limit (MDL). In this file, these 'non-detects' are reported as zero and the QACODE is set to "CHM-A" to indicate the assignment. While the concentration of the analyte is clearly small, it is not strictly zero. The MDL is therefore listed as a guideline to users who wish to substitute values other than zero, i.e., setting the non-detect value to the MDL value, half the MDL value, etc. Furthermore, results of organic analytes may routinely show non-zero values that are less than the MDL. This apparent inconsistency is possible because, by convention, the MDLs for organic analyses are calculated to indicate the threshold of reliable measurements, rather than the stricter limit of instrumental detection. In these cases, the best estimate of the concentration is reported (i.e., the value reported by the analytical laboratory), the QACODE is set to "CHM-B", and the MDL is listed. The user can be confident that the analyte is present, but there is a high degree of uncertainty in the reported concentration. Note that the value of the MDL depends on the

dilution history of the sample; therefore, its magnitude can differ widely among samples. Most results in this file are larger than the MDL and are reported directly without MDL values or QACODEs. Finally, records flagged with "CHM-C" indicate that the concentration value is uncertain because an interference was noted in the blank analysis performed with the sample; caution is advised in interpreting these results. To summarize:

QACODE	INTERPRETATION	CONC reported	MDL reported
<none>	result is detectable and > MDL	as measured	<none>
CHM-A	result is ≤ MDL and undetectable	zero	MDL is listed
CHM-B	result is ≤ MDL but detectable	best estimate	MDL is listed
CHM-C	result may be affected by interference	best estimate	<none>

Three analytical labs were involved in analyzing sediment analytes in 2000 and 2001: two state labs for sediment samples collected by co-operative teams in Connecticut and New York (designated by LABCODE = NY and CT) and a national contract lab for samples collected in other northeastern states (LABCODE = NAT). In two respects, there are noticeable differences in results attributable to different methods used by the three labs. (1) There is a distinction in the number of "non-detects" (concentrations less than the method detection limit or MDL) evident among labs, probably arising from different MDL values used by the labs during analysis. (2) The labs used different procedures to digest sediment samples prior to metal analysis, affecting results for several metals. Refer to Section 5.2.6 for discussion of the nature and implications of the discrepancies.

NCA planners provide two alternate locations for a station location in the event that the original location cannot be sampled. The parameter STA_ALT indicates whether the station location was the original site, first alternate, or second alternate—STA_ALT = "A", "B", or "C", respectively. Also refer to discussion in the STATIONS metadata file regarding use of this parameter during analysis of the data.

4.4 Summary of Dataset Parameters

* denotes parameters that should be used as key fields when merging data files

*STATION	Station name
*STAT_ALT	Alternate Site Code (A, B, C)
*EVNTDATE	Event date
*ANALYTE	Name of analyte measured. A list of the ANALYTE codes and their full chemical names is presented in the file ANALYTES; also see Section 7.1.3.
CONC	Concentration of analyte. Results fall into one of three categories: 1) the analyte concentration was large and reliably reported; 2) the analyte concentration was less than the method detection limit, but the best estimate of the concentration is reported; and 3) and the analyte was not detected and is reported as zero. See Section 4.3 for further discussion.
CHMUNITS	Concentration units used to report results, reported as the

mass of analyte per dry mass of sediment:
Metals ug/g (ppm)
PAHs, PCBs, Pesticides ng/g (ppb)

MDL — Method Detection Limit; reported only when measured concentration is < MDL (see Section 4.3)

QACODE — QA/QC codes:

- <blank> CONC > MDL; concentration value is reliable
- CHM-A CONC is undetectable; value set to zero (user may wish to substitute another value)
- CHM-B CONC ≤ MDL, but is detectable; best estimate reported
- CHM-C failed QA criteria: an interference was noted in the blank analysis performed with the sample; caution is advised in interpreting the result

See Section 4.3 for further discussion.

LABCODE — Code identifying laboratory responsible for performing chemical analyses

- CT State laboratory for CT samples
- NY State laboratory for NY samples
- NAT National contract lab for other Northeast states

5.0 DATA ACQUISITION AND PROCESSING METHODS

5.1 Data Acquisition / Field Sampling

The sample collection methods used by USEPA trained field crews will be described here. Any significant variations by NCA partners are noted in Section 5.1.12. Details regarding NCA partners are reported in the STATIONS data file.

5.1.1 Sampling Objective

Sediment sub-samples were collected for the analysis of metallic and organic chemical constituents. Separate sub-samples from the same grab were used for sediment grain-size analyses and toxicity testing. Additional sediment grabs were taken for benthic macrofaunal analysis.

5.1.2 Sample Collection: Methods Summary

Sediment was collected with a 0.04-m^2 Young-modified Van-Veen grab or similar sampler. Only the top two centimeters of a grab were retained for physical, chemical, and toxicological analyses. A sufficient number of grabs were processed to provide three liters of the 2-cm composite material. The composite was homogenized and separated into two fractions for storage until analysis. One fraction was frozen and used in the measurement of total organic carbon (TOC) and concentrations of chemical contaminants. The second fraction was chilled but not frozen during storage, and was used for grain-size and toxicity analyses. Separate sediment grabs were taken for benthic macrofaunal analysis.

5.1.3 Beginning Sampling Dates

25 June 2001

5.1.4 Ending Sampling Dates
31 October 2001

5.1.5 Sampling Platform
Samples were collected from gasoline or diesel powered boats, 18 to 133 feet in length.

5.1.6 Sampling Equipment
A 1/25 m2, stainless steel (coated with Kynar), Young-modified Van Veen grab sampler was used to collect sediments.

5.1.7 Manufacturer of Sampling Equipment
Young's Welding, Sandwich, MA

5.1.8 Key Variables
Not applicable

5.1.9 Sample Collection: Methods Calibration
The sampling gear does not require calibration, although it was inspected regularly for damage by mishandling or impact on rocky substrates.

5.1.10 Sample Collection: Quality Control
Care was taken to minimize disturbance to the sediment grabs. Grabs that were incomplete, slumped, less than 7 cm in depth, or comprised chiefly of shelly substrates were discarded. The chance of sampling the same location was minimized by repositioning the boat five meters downstream after three sampling attempts.

5.1.11 Sample Collection: References
Strobel, C.J. 2000. Environmental Monitoring and Assessment Program: Coastal 2000 - Northeast component: field operations manual. Narragansett (RI): U.S. Environmental Protection Agency, National Health and Environmental Effects Research Laboratory, Atlantic Ecology Division. Report nr EPA/620/R-00/002. 68 p.

5.1.12 Sample Collection: Alternate Methods
Different grab samplers used by NCA partners include the Smith-MacIntyre and Ponar grab samplers.

5.2 Data Preparation and Sample Processing

5.2.1 Sample Processing Objective
Sediment samples were analyzed for total metals, PAHs, PCBs and pesticides.

5.2.2 Sample Processing: Methods Summary
All analyses were performed on samples that were stored frozen. Sediments analyzed for total metals were dried and completely digested in nitric/hydrofluoric acids (acid persulfate for mercury). The analytical methods used to measure analyte concentrations were: cold vapor atomic analysis (AA) for mercury; graphite furnace AA for silver,

arsenic, cadmium, lead, antimony, tin and thallium; hydride generation atomic fluorescence for selenium; and optical-emission ionically coupled plasma (ICP) for the remaining metals. For the organic analyses, sediments were extracted using the procedures of NOAA National Status and Trends Program (Lauenstein and Cantillo, 1993). The PAHs were analyzed by gas-chromatography / mass-spectrometry (GC/MS); pesticides and PCBs were analyzed by GC/ECD (electron capture detector).

5.2.3 Sample Processing: Calibration

The analytical instruments were calibrated by standard laboratory procedures including: constructing calibration curves, running blank and spiked quality control samples, and analyzing standard reference materials.

5.2.4 Sample Processing: Quality Control (QC)

Each batch of samples was accompanied by QC analyses consisting of method blanks, matrix spikes, matrix spike duplicates, and standard reference materials (SRMs). In total, approximately 5% of all analyses were QC analyses. Processing quality was considered acceptable if the following criteria were met: blanks were less than three times the minimum detection limit; accuracy, as determined by analysis of certified reference materials, was within 30% for organic analytes and within 15% for inorganic analytes; and precision, as determined by replicate analyses, was within 30% for organic analytes and within 15% for inorganic analytes. Additional specifications and guidelines are presented in U.S. EPA 2001.

5.2.5 Sample Processing: References

Lauenstein, G. G. and A. Y. Cantillo (eds.). 1993. Sampling and analytical methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992: Comprehensive descriptions of trace organic analytical methods, Volume IV NOAA Technical Memorandum NOS ORCA 71, Silver Spring, MD. 182 pp.

Texas A & M University, Geochemical and Environmental Research Group. 1990. NOAA Status and Trends, Mussel Watch Program, Analytical Methods. Submitted to NOAA. Rockville (MD): U.S. Dept. of Commerce, National Oceanic & Atmospheric Administration, Ocean Assessment Division.

U.S. EPA. 1995. Environmental Monitoring and Assessment Program (EMAP): Laboratory Methods Manual-Estuaries, Volume 1: Biological and Physical Analyses. Narragansett (RI): U.S. Environmental Protection Agency, Office of Research and Development, EPA/620/R-95/008.

U.S. EPA. 2001. Environmental Monitoring and Assessment Program (EMAP): National Coastal Assessment Quality Assurance Project Plan 2001-2004. U.S. Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002. 189 p

5.2.6 Sample Processing: Alternate Methods

Three analytical labs were involved in analyzing sediment analytes in 2000 and 2001: two state labs for sediment samples collected by co-operative teams in Connecticut and New York (designated by LABCODE = NY and CT) and a national contract lab for samples collected in other northeastern states (LABCODE = NAT). In two respects, there are noticeable differences in results attributable to different methods used by the three labs. (1) There is a distinction in the number of "non-detects" (concentrations less than the method detection limit or MDL) evident among labs, probably arising from different MDL values used by the labs during analysis. (2) The labs used different procedures to digest sediment samples prior to metal analysis, affecting results for several metals.

Different incidence of non-detects reported by analytical labs. Listed below are the percentages of records that are non-detects, distinguished by year, LABCODE, and type of analyte: pesticide, PCBs, PAHs, and metals. Non-detects are reported as zero in the NCA database (see Section 4.3).

Percentage of non-detects (zeros) in NCA 2000 & 2001 SEDCHEM file.

YEAR	LABCODE	pesticide	PCB	PAH	metal
2000	CT	93	81	53	8
	NY	88	66	0	18
	NAT	65	31	4	5
		pest	PCB	PAH	metal
2001	CT	98	95	59	8
	NY	86	64	0	23
	NAT	64	31	1	4

Note the following: 1) A relatively large percentage of pesticide analyses are non-detects (an acceptable situation). However, there is a consistent difference in the 2000 & 2001 pesticide data among labs: CT > NY > NAT. Also, almost all CT analyses in 2001 were non-detects, a larger fraction than for CT in 2000. 2) The same observations hold for PCBs, i.e., CT > NY > NAT and CT2001 > CT2000. 3) For PAHs, only CT had significant incidence of non-detects in either year. 4) Relatively few non-detects were reported for metals, with roughly equal incidence among labs and years.

The three analytical labs used different MDL values when measuring chemical concentrations in sediment. In the case of pesticides and PCBs, the average MDL values were approximately 1 ppb for NY; 0.5 ppb for CT, and 0.25 ppb for NAT (averaged for all pesticides or PCBs). This observation may explain the relatively small rate of non-detects in pesticide and PCB analyses performed by the national contract; however, it doesn't explain the relative rate distinction between the NY and CT labs. Similarly, MDL values differed for PAH analyses: about 10 ppb for CT and about 2 ppb for NAT (NY did not report any MDLs for PAHs). MDL values were similar for all labs regarding metal analyses. Generally,

the MDLs used by all labs were smaller than the limits required by the NCA program. In short, much of the discrepancy noted in the percentages of non-detects in 2000 & 2001 data probably arose because of the different MDL values employed by the three labs.

The implications of the differences highlighted above depend on how the data are used. Generally, the non-detects may be interpreted as 'very small concentrations'. Thus, the number of non-detects may be immaterial, for example, if the data are used primarily to identify polluted sites. However, the number of non-detects (zero values) may significantly affect calculated metrics such as averages, medians, expressions of variability, etc. Year-to-year inconsistencies in the use of MDL values may also confound the interpretation of temporal trends.

Different sediment digestion methods. Another important difference is evident among the three laboratories regarding the method of digestion performed on sediments prior to metal analysis in both 2000 and 2001. The national contract lab (LABCODE = NAT) used an HF/HNO3 digestion, while the state labs (LABCODE = CT and NY) originally used a less aggressive HNO3 digestion. The CT lab re-analyzed a subset of metals analytes (Aluminum, Iron, Manganese, Lead Nickel Cadmium, Silver, Selenium, Animony) using FG/HN03 digestion. All other CT metals and all the NY metals were measured using HN03 digestion. To investigate the likely effects of the differing methods, archived sediment from all NY samples collected in 2000 were reanalyzed using the more aggressive HF/HNO3 digestion. A comparison of results can be expressed as linear regression of HF/HNO3 results vs HNO3 results:

Linear regression parameters of concentrations measured following an HF/HNO3 digestion vs an HNO3 digestion, i.e., HF/HNO3 = m*HNO3 + b:

Metal	slope (m)	intercept (b)	R^2
Al	3.13	15300	0.74
Fe	0.98	4660	0.92
Mn	0.76	203	0.73
As	0.68	0.47	0.78
Pb	0.95	9.70	0.95
Hg	1.01	-0.02	0.85
Ni	1.20	0.45	0.92
Zn	0.94	11.1	0.97
Cd	0.94	-0.68	0.77
Cr	1.41	10.5	0.96
Cu	1.33	-1.97	0.89
Ag	0.68	-0.6	0.81
Se	0.72	-0.28	0.44
Sb	0.02	0.08	0.09

Perfect agreement of methods would result in parameter values of m = 1, b = 0, and R^2 = 1. The HF/HNO3 digestion completely dissolves the aluminosilicate matrix of sediments, yielding significantly larger concentrations of crustal elements: Al, Fe, and Mn. Relatively good agreement was evident for both digestion methods for the most toxic

elements that have designated ERM (effects range median) limits: As, Pb, Hg, Ni, Zn, Cd, Cr, and Cu (Pb and Cr values may be marginally elevated in HF/HNO3 digestions). There was poor agreement for the non-priority elements selenium (Se) and antimony (Sb). Note that the NY data included in this database are the original results obtained following the non-standard HNO3 digestion. All data for the inter-comparison study of the 2000 NY sediments are included in an Excel file titled "NY metals compare" available from the NCA website.

6. DATA ANALYSIS AND MANIPULATIONS

6.1 Name of New or Modified Values

Not applicable

6.2 Data Manipulation Description

Concentrations of metallic analytes smaller than the method detection limit were reported as zero (see Section 4.3 for details).

7. DATA DESCRIPTION

7.1 Description of Parameters

7.1.1 Components of the Dataset

PARAMETER	TYPE	LENGTH	LABEL
ANALYTE	Char	8	Code for Analyte Measured
CONC	Num	8	Concentration of Analyte in Sample
CHMUNITS	Char	10	Unit of Measure
MDL	Num	8	Method Detection Limit
STATION	Char	9	Station Name
STAT_ALT	Char	1	Station Name
EVNTDATE	Num	8	Event Date
QACODE	Char	10	QA Code(s)
LABCODE	Char	8	Contract/Lab Identifier

7.1.2 Precision of Reported Values

All values have been rounded to three significant digits.

7.1.3 Minimum and Maximum Value in Dataset (non-zero data)

* Comments indicate exceptions by analytical labs (LABCODE)
Blank entries in MIN and MAX columns indicate non-detects

ANALYTE ID	ANALYTE NAME	MIN	MAX	Comment*
Metals				
AG	Silver	0.06	6.95	
AL	Aluminum	167	78600	

AS	Arsenic	1	88.1	
CD	Cadmium	0.02	40.6	
CR	Chromium	2	332	
CU	Copper	1	657	
FE	Iron	4.41	53100	
HG	Mercury	0.01	2.64	
MN	Manganese	24	1790	
NI	Nickel	1	55	
PB	Lead	0.306	278	
SB	Antimony	0.1	28.3	
SE	Selenium	0.05	40.8	
SN	Tin	0.1	11900	except NY
ZN	Zinc	4	780	

Polynuclear aromatic hydrocarbons (PAHs)

ACENTHE	Acenaphthene	0.031	490	
ACENTHY	Acenaphthlylene	0.03	560	
ANTHRA	Anthracene	0.042	3500	
BENANTH	Benz(a)anthracene	0.35	5800	
BENAPY	Benz(a)pyrene	0.085	5200	
BENEPY	Benz(e)pyrene	7.89	321	CT only
BENZOBFL	Benzo(b)fluoranthene	0.069	5400	
BENZOKFL	Benzo(k)fluoranthene	0.03	1400	
BENZOP	Benzo(g,h,i)perylene	0.056	2200	
BIPHENYL	Biphenyl	0.044	860	
CHRYSENE	Chrysene	0.089	4600	
DIBENTP	Dibenzothiophene	0.03	590	
DIBENZ	Dibenz(a,h)anthracene	0.052	800	
DIMETH	2,6-dimethylnaphthalene	0.039	250	
FLUORANT	Fluoranthene	0.23	10000	
FLUORENE	Fluorene	0.074	710	
INDENO	Indeno(1,2,3-c,d)pyrene	0.076	2400	
MENAP1	1-methylnaphthalene	0.065	190	
MENAP2	2-methylnaphthalene	0.094	150	
MEPHEN1	1-methylphenanthrene	0.038	1200	
METH	Methoxychlor			CT only
NAPH	Naphthalene	0.27	410	
PHENANTH	Phenanthrene	10.4	1200	NAT only
PYRENE	Pyrene	0.13	8500	
TRIMETH	2,3,5-trimethylnaphthalene	0.025	230	

Polychlorinated biphenyls (PCBs)

PCB101	2,2',4,5,5'-pentachlorobiphenyl	0.01	580	
PCB105	2,3,3',4,4'-pentachlorobiphenyl	0.009	200	
PCB110	2,2',4,5,5'pentachlorobiphenyl	1	41	NY only
PCB118	2,3',4,4',5-pentachlorobiphenyl	0.012	690	
PCB126	3,3',4,4',5-pentachlorobiphenyl	0.15	0.36	
PCB128	2,2',3,3',4,4'-hexachlorobiphenyl	0.007	110	
PCB138	2,2',3,4,4',5'-hexachlorobiphenyl	0.016	510	
PCB153	2,2',4,4',5,5'-hexachlorobiphenyl	0.015	590	

PCB170	2,2',3,3',4,4',5-heptachlorobiphenyl	0.028	42	
PCB18	2,2',5-trichlorobiphenyl	0.029	371	
PCB180	2,2',3,4,4',5,5'-heptachlorobiphenyl	0.017	84	
PCB187	2,2',3,4',5,5',6-heptachlorobiphenyl	0.007	44	
PCB195	2,2',3,3',4,4',5,6-octachlorobiphenyl	0.003	8.5	
PCB206	2,2',3,3',4,4',5,5',6-nonachlorobiphenyl	0.005	30	
PCB209	decachlorobiphenyl	0.003	48	
PCB28	2,4,4'-trichlorobiphenyl	0.021	900	
PCB44	2,2',3,5'-tetrachlorobiphenyl	0.018	260	
PCB52	2,2',5,5'-tetrachlorobiphenyl	0.011	550	
PCB66	2,3',4,4'-tetrachlorobiphenyl	0.011	367	
PCB77	3,3',4,4'-tetrachlorobiphenyl	0.032	25	
PCB8	2,4'-dichlorobiphenyl	0.021	82	
Pesticides				
ABHC	alpha-Hexachlorohexane	31.9	31.9	CT only
ALDRIN	Aldrin	0.28	12	
BBHC	beta-Hexachlorohexane			CT only
CISCHL	alpha-Chlordane	0.006	8.9	except NY
CNONCHL	cis-Nonachlor			CT only
DBHC	delta-Hexachlorohexane	6.77	6.77	CT only
DIELDRIN	Dieldrin	0.007	26	
ENDOSUI	Endosulfan I	0.036	9.38	
ENDOSUII	Endosulfan	0.023	9.5	
ENDOSULF	Endosulfan II	0.068	4.7	
ENDRIN	Endrin	0.034	0.25	
ENDRINA	Endrin-a			CT only
ENDRINK	Endrin-k			CT only
GBHC	gamma-Hexachlorohexane	1.17	130	CT only
HEPTACHL	Heptachlor	0.012	2.7	
HEPTAEPO	Heptachlor epoxide	0.013	3	
HEXACHL	Hexachlorobenzene	0.002	13	
LINDANE	Lindane (gamma-BHC)	0.005	3	
MIREX	Mirex	0.005	9	
OPDDD	2,4'-DDD	0.012	120	
OPDDE	2,4'-DDE	0.014	71	
OPDDT	2,4'-DDT	0.011	2.4	except NY
OXYCHL	Oxychlordane			CT only
PPDDD	4,4'-DDD	0.015	24	except NY
PPDDE	4,4'-DDE	0.005	88	
PPDDT	4,4'-DDT	0.009	250	
TNONCHL	trans-Nonachlor	0.005	12	
TOXAPHEN	Toxaphene			

7.1.4 Maximum Value in Dataset

See Section 7.1.3

7.2 Data Record Example

7.2.1 Column Names for Example Records

STATION	STAT_ALT	EVNTDATE	ANALYTE	CONC	QACODE	MDL	CHMUNITS	LABCODE

7.2.2 Example Data Records

STATION	STAT_ALT	EVNTDATE	ANALYTE	CONC	QACODE	MDL	CHMUNITS	LABCODE
CT01-0001	A	8/17/00	ABHC	0	CHM-A	0.416	ng/g	CT
CT01-0001	A	8/17/00	ACENTHE	0	CHM-A	8.33	ng/g	CT
CT01-0001	A	8/17/00	ACENTHY	0	CHM-A	8.33	ng/g	CT
CT01-0001	A	8/17/00	AG	0.75		.	ug/g	CT
CT01-0001	A	8/17/00	AL	4780	CHM-C	1500	ug/g	CT

8. GEOGRAPHIC AND SPATIAL INFORMATION

8.1 Minimum Longitude (Westernmost)
-75.6977 decimal degrees

8.2 Maximum Longitude (Easternmost)
-67.0482 decimal degrees

8.3 Minimum Latitude (Southernmost)
38.4739 decimal degrees

8.4 Maximum Latitude (Northernmost)
45.1848 decimal degrees

8.5 Name of Region
The National Coastal Assessment Northeast Region covers the northeastern US coastline from Maine to Delaware

9. QUALITY CONTROL AND QUALITY ASSURANCE

9.1 Measurement Quality Objectives
Measure replicate grain size of samples to within a precision of 10% (see U.S. EPA 2001).

9.2 Data Quality Assurance Procedures

9.3 Actual Measurement Quality

10. DATA ACCESS

10.1 Data Access Procedures
Data can be downloaded from the web
http://www.epa.gov/emap/nca/html/regions/index.html

10.2 Data Access Restrictions
None

10.3 Data Access Contact Persons

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401-782-3034, 401-782-3030 (FAX), kiddon.john@epa.gov

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10.4 Dataset Format
ASCII (CSV) and SAS Export files

10.5 Information Concerning Anonymous FTP
Not available

10.6 Information Concerning WWW
No gopher access, see Section 10.1 for WWW access

10.7 EMAP CD-ROM Containing the Dataset
Data not available on CD-ROM

11. REFERENCES

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12. TABLE OF ACRONYMS

AED	Atlantic Ecology Division
CSC	Computer Sciences Corporation
EMAP	Environmental Monitoring and Assessment Program
EPA	Environmental Protection Agency
MDL	Method Detection Limit
NCA	National Coastal Assessment
ng/g	Nano gram per gram
NHEERL	National Health and Environmental Effects Research Laboratory
PAH	Polynuclear Aromatic Hydrocarbon
PCB	Polychlorinated Biphenyls
ppb	parts per billion
ppm	parts per million
QA/QC	Quality Assurance/Quality Control
SRM	Standard Reference Material
TOC	Total Organic Carbon
ug/g	Micro gram per gram
WWW	World Wide Web

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